Study Title
Antibacterial Activity and Efficacy Evaluation of UVC Cleaning System's UV Device

Test Method
Custom Device Study Based on: ASTM E1153
Efficacy of Sanitizers Recommended for Inanimate Non-Food Contact Surfaces

Study Identification Number
NG5622-III

Study Sponsor
UVC Cleaning Systems
7876 S Van Dyke Rd
Marlette, MI 48453

Test Facility
Antimicrobial Test Laboratories
1304 W. Industrial Blvd
Round Rock, TX 78681
(512) 310-8378
History of the Laboratory

Antimicrobial Test Laboratories was launched in 2006 to provide testing services to the antimicrobial industry. The company has grown considerably since then but its focus remains the same: Test antimicrobial agents, test them well, and test them fast! Antimicrobial Test Laboratories operates a 15,000+ square foot facility near Austin, Texas, where hundreds of studies are conducted annually by a staff of friendly, knowledgeable, and experienced microbiologists and virologists.

Laboratory Qualification Statement

Antimicrobial Test Laboratories was founded by microbiologist Dr. Benjamin Tanner. The laboratory ensures consistent, reproducible results by utilizing a well-trained and educated scientific staff who work from a comprehensive system of Standard Operating Procedures, official standard methods from ASTM, AOAC, and other organizations, and custom study protocols. The laboratory provides testing services to dozens of Fortune 500 companies and has been inspected for GLP compliance by the US government.

Scientist Qualifications

This study was designed, conducted, and reported by: Katelyn Hammond, B.S.

Katelyn graduated from the University of Texas with a Bachelors of Science in Microbiology.

Katelyn is well-versed with regard to a variety of microbiological test methods and procedures. As a Microbiologist at Antimicrobial Test Laboratories, she has taken part in hundreds of studies and mastered several test methods. Katelyn works with clients throughout the course of their projects to ensure that their technical needs are met. She is highly regarded in the laboratory for her keen troubleshooting skills and positive attitude.

If you have any questions about your study, please don't hesitate to contact Katelyn at:

Katelyn@AntimicrobialTestLabs.com
or
(512) 310-8378
Test Device Information

Test Device: M15
Setup and operated by Study Sponsor, 17-19NOV2014.

Test Microorganism Information

The test microorganism(s) selected for this test:

Listeria monocytogenes
This bacteria is a Gram-positive, rod shaped, facultative anaerobe that is motile due to the presence of flagella. These bacteria are common cause of the foodbourne illness listeriosis, which can be fatal. Listeriosis can cause meningitis and sepsis and is particularly dangerous to pregnant women and unborn infants. Listeria monocytogenes is pervasive and can be found in soil, water, and certain livestock animals. They can resist both heat and freezing and can survive for several years.
Summary of the Procedure

- An overnight culture was centrifuged at 1,000 RPM (as applicable) for 10 minutes, re-suspended in sterile R/O water, and supplemented with 5% FBS.
- Stainless steel carriers (1”x3”) were inoculated with 0.01 ml of the culture.
- Inoculum was spread over approximately 10 cm$^2$ of each carrier.
- Carriers were left to dry at room temperature for 10 minutes in Petri dishes with lids ajar.
- Visibly dry carriers/test microorganism were harvested in 20 ml D/E to determine the initial numbers control.
- Test carriers were treated at a specified distance and time period then harvested in 20 ml D/E.
- Standard dilution and pour plating techniques were used for all enumerations.
- Calculations are based off of the averaged initial and final control numbers compared to CFU/carrier recovered from treated test carriers.

Study Timeline

<table>
<thead>
<tr>
<th>Culture Inoculated</th>
<th>Carriers Inoculated</th>
<th>Carriers Treated</th>
<th>Carriers Harvested</th>
<th>Carriers Evaluated</th>
<th>Report Delivered</th>
</tr>
</thead>
</table>

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Criteria for Scientific Defensibility of a Custom Device Study

For Antimicrobial Test Laboratories to consider a Device Study study to be scientifically defensible, the following criteria must be met:

1. The average number of viable bacteria recovered from the time zero samples must be approximately $1 \times 10^5$ cells/carrier or greater.
2. Positive/Growth controls must demonstrate growth of the appropriate test microorganism.
3. Negative/Purity controls must demonstrate no growth of test microorganism.

Passing Criteria

Because of the nature of the study, passing criteria may be determined by the Study Sponsor.

Testing Parameters used in this Study

- **Carrier (Size):** Stainless Steel (1” x 3”)
- **Replicates:** See Data
- **Culture Dilution Media:** Sterile Reverse Osmosis (R/O) Water
- **Culture Supplement:** 5% FBS
- **Inoculum Target:** $1.0 \times 10^7$ CFU/Carrier
- **Inoculum Volume:** 0.01 ml
- **Inoculum Surface Area:** 10 cm$^2$
- **Carrier Dry Time:** 10 Minutes
- **Carrier Dry Temperature:** Ambient (23±2°C)
- **Contact Time(s):** See Data
- **Contact Distance(s):** See Data
- **Contact Temperature:** Ambient (23±2°C)
- **Neutralizer (Vol.):** D/E Broth (20 ml)
- **Enumeration Media, Method:** TSA, pour plate
- **Enum. Media Supplement:** N/A
- **Enum. Plate Incubation Temp.:** 36°C ± 1°C
- **Enum. Plate Incubation Time:** 24-48 hours
- **Enum. Plate Incubation Conditions:** Aerobic
Study Notes

Test Room Dimensions: 11’ 3” x 18’ 8” (8’ ceiling)

Study Photographs

Top: Inoculated carrier drying at room temperature
Bottom: Test carriers at 5 meters and 5 meters, perpendicular prior to treatment.
Control Results

Neutralization Method: D/E (20 ml) Vortex  
Growth Confirmation: Colony Morphology  
Media Sterility: Sterile  
Antibiotic Resist: N/A

Calculations

\[
\text{Percent Reduction} = \left( \frac{B - A}{B} \right) \times 100
\]

Where:
B = Average number of viable test microorganisms on the control carriers  
A = Number of viable test microorganisms on the test carriers after the contact time

\[
\log_{10} \text{Reduction} = \log\left( \frac{B}{A} \right)
\]

Where:
B = Average number of viable test microorganisms on the control carriers.  
A = Number of viable test microorganisms on the test carriers after the contact time
### Results

<table>
<thead>
<tr>
<th>Test Microorganism</th>
<th>Device</th>
<th>Contact Time</th>
<th>Contact Distance</th>
<th>CFU/Carrier</th>
<th>Geometric Mean CFU/Carrier</th>
<th>% Reduction vs Numbers Control</th>
<th>Log_{10} Reduction vs Numbers Control</th>
</tr>
</thead>
<tbody>
<tr>
<td>L. monocytogenes ATCC 15313 (Run 1)</td>
<td>M15</td>
<td>Numbers Control</td>
<td>8.50E+05</td>
<td>4.11E+05</td>
<td>N/A</td>
<td></td>
<td></td>
</tr>
<tr>
<td></td>
<td></td>
<td>30 Minutes</td>
<td>1.99E+05</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td></td>
<td></td>
<td>5 Meters</td>
<td>&lt;1.00E+00</td>
<td>&lt;1.00E+00</td>
<td>&gt;99.9998%</td>
<td>&gt;5.61</td>
<td></td>
</tr>
<tr>
<td></td>
<td></td>
<td>5 Meters, Perpendicular</td>
<td>&lt;1.00E+00</td>
<td>&lt;1.00E+00</td>
<td>&gt;99.9998%</td>
<td>&gt;5.61</td>
<td></td>
</tr>
</tbody>
</table>

Note: The limit of detection for this study is 1.00E+00 CFU/Carrier. Values below this limit are shown as 0 in the chart above.

The results of this study apply to the tested substances(s) only. Extrapolation of findings to related materials is the responsibility of the Sponsor.

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